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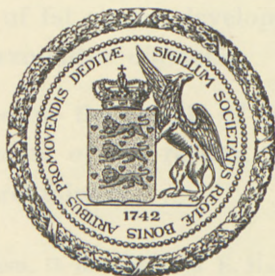
SOME
STUDIES ON VITAMIN E

BY

H. DAM, J. GLAVIND, INGE PRANGE

AND

J. OTTESEN



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I KOMMISSION HOS EJNAR MUNKSGAARD

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Previous communications from this laboratory have dealt with two nutritional diseases in chicks, viz. the previously known alimentary encephalomalacia¹ and the new disease alimentary exudative diathesis². It was shown that these diseases are due to a deficiency of vitamin E.

In this paper, our continued studies on vitamin E are reported. The studies can be divided into the following four groups:

1. The chemical determination of tocopherols.
2. Attempts to use the exudative diathesis for the comparison of the activity of natural sources of vitamin E with α -tocopherol.
3. Comparison between the anti-exudative activity of α - and β -tocopherol.
4. The influence of fat on the development of encephalomalacia and exudative diathesis.

This paper presents the results of our studies on these subjects, though some of the results, however, are to be looked upon as preliminary.

¹ H. DAM, J. GLAVIND, O. BERNTH and E. HAGENS, *Nature* **142**, 1157 (1938).

² H. DAM and J. GLAVIND, *Nature* **142**, 1077, (1938). H. DAM and J. GLAVIND, *Nature* **143**, 810 (1939). H. DAM and J. GLAVIND, *Skand. Archiv für Physiol.* **82**, 299 (1939). H. DAM and J. GLAVIND, *Die Naturwissenschaften* **28**, 207 (1940).

I. The chemical determination of tocopherols.

For the chemical determination of vitamin E (total amount of tocopherols), several methods are described which we have tried to apply and to improve.

1. The spectrographic method. The use of the absorption spectrum of vitamin E for the chemical determination has been proposed. CUTHBERTSON et al.¹ used the method for body fat and found that tocopherol could be estimated with reasonable accuracy in crude extract when present in fat in amounts down to 1 %, and in the unsaponifiable fraction when present in amounts down to 0,1 %. A modification of the method consisting in oxidation of the unsaponifiable fraction with silver nitrate made a detection of 0,01 % possible.

We have tried to use the method for pure d,l- α -tocopherol, for wheat germ oil, and for the unsaponifiable fraction of wheat germ oil. The results appear from Fig. 1 which shows the absorption spectra measured by means of a Hilger quartz spectrograph with echelon cells.

Curve 1 shows the absorption spectrum of pure d,l- α -tocopherol in absolute alcohol with a maximum at 293 m μ . Curve 2 gives the absorption spectrum of wheat germ oil in hexane; no maximum at 293 m μ . Curve 3 represents the absorption spectrum of unsaponifiable matter from wheat germ oil in absolute methyl alcohol; it shows a distinct maximum at 293 m μ . The extinction is, however, about 50 % greater than that corresponding to the tocopherol content as determined by the method of EMMERIE and ENGEL. Unsaponifiable matter of extract of alfalfa, even after distillation in a high vacuum, showed no

¹ W. F. J. CUTHBERTSON, R. R. RIDGEWAY and J. C. DRUMMOND, *Biochem. J.* **34**, 34 (1940).

maximum at 293 $m\mu$, but a distinct inflection. However, the extinction was only to about 20 % due to tocopherols.

For an exact spectrographic determination of tocopherols, a further purification of the extracts, which is rather difficult, will therefore be necessary. Even if an extinction

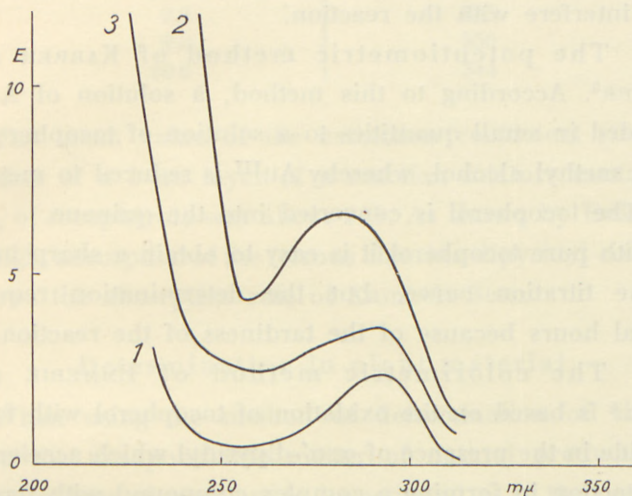


Fig. 1. Abscissae: wave length in $m\mu$.

Ordinates: extinction.

Curve 1: d,l- α -tocopherol 0.032 %.

Curve 2: Wheat germ oil in hexane 10.7 %, corresponding to 0.026 % of tocopherol in the solution.

Curve 3: Unsaponifiable matter from 1.299 g wheat germ oil in 10 cc methyl alcohol corresponding to 0.032 % of tocopherol in the solution.

four times as great is obtained by oxidation with $AgNO_3$, as used by CUTHBERTSON et al., this will not suffice for an exact tocopherol determination in plant material.

2. The colorimetric method of FURTER and MEYER¹. By means of this method, the red colour developed after heating an alcoholic solution of tocopherol with nitric acid is estimated. We found, however, that this method

¹ M. FURTER and R. E. MEYER, *Helv. chim. acta* **22**, 240 (1939).

was not exact, and the results were more difficult to reproduce than when the method of EMMERIE and ENGEL was applied; therefore we worked exclusively with the last mentioned method. Other authors have found that catechol derivatives¹ and the quinone corresponding to α -tocopherol² may interfere with the reaction.

3. The potentiometric method of KARRER and KELLER³. According to this method, a solution of AuCl_3 is added in small quantities to a solution of tocopherol in 80 % methyl alcohol, whereby Au^{III} is reduced to metallic Au. The tocopherol is converted into the quinone.

With pure tocopherol it is easy to obtain a sharp break in the titration curve, but the determination requires several hours because of the tardiness of the reaction.

4. The colorimetric method of EMMERIE and ENGEL⁴ is based on the oxidation of tocopherol with ferric chloride in the presence of α : α' -dipyridyl which accelerates the reaction in forming a complex compound with ferrous ions. This complex has a red colour which may be measured after 15 min. by filter S 50 in the Zeiss-Pulfrich photometer. The reaction is carried out in ethyl alcohol which may contain up to 20 % of benzene.

Determination of the extinction coefficient.

With pure α -tocopherol the method works fairly well. We prepared the α -tocopherol by saponifying the pure acetate and used these solutions for the determination of the extinction coefficient (Table 1).

¹ L. I. SMITH, W. B. IRWIN and H. F. UNGNADE, *J. Am. Chem. Soc.* **61**, 2424 (1939).

² A. EMMERIE and C. ENGEL, *Rec. trav. chim.* **58**, 284 (1939).

³ P. KARRER and H. KELLER, *Helv. chim. acta* **21**, 1161 (1938).

⁴ A. EMMERIE and C. ENGEL, *Rec. trav. chim.* **57**, 1351 (1938).

Table 1

mg d,l- α -tocopherol acetate saponified	Extinction coefficient E $\frac{1 \text{ cm}}{1\%}$
268.3	345
9.8	335
9.8	350
10.6	344

The mean value of the extinction coefficient (the extinction of a 1 cm layer of a reaction mixture containing 1 % of tocopherol) was about 345. As shown by EMMERIE, the light absorption of the ferrous chloride-dipyridyl-complex follows the absorption rule of Lambert-Beer.

Determination in plant material.

When using the method of determination of vitamin E in the unsaponifiable matter of natural products, the same sources of error are to be expected as in the potentiometric method.

EMMERIE and ENGEL¹ proposed to remove carotenoids by adsorption on columns of flordin XS earth. The carotenoids are said to be adsorbed while tocopherol is passing into the filtrate. We were not able to obtain reliable results in this way.

A more suitable method to eliminate carotenoids and certain other reducing substances is the acetylation method developed by KARRER and KELLER² in their potentiometric method. They heated a mixture of β -carotene and tocopherol with acetic anhydride in pyridine solution on the water bath for two hours. The pyridine solution is then

¹ A. EMMERIE and C. ENGEL, *Rec. trav. chim.* **58**, 3 (1939).

² P. KARRER and H. KELLER, *Helv. chim. acta* **22**, 253 (1939).

diluted with petrol ether, washed with diluted hydrochloric acid, and then with water until neutral reaction. Then the petrol ether is evaporated and the remaining substances are titrated with auric chloride. Another sample of the carotene-tocopherol-mixture is titrated without acetylation and the tocopherol content is evaluated as the difference between the two titrations. KARRER and KELLER found that by this method a fairly good separation between tocopherol and carotene is obtained. We carried out similar experiments with mixtures of carotene and tocopherol and carotene alone, but we made the determinations by means of the colorimetric method. We were able to state the results of KARRER and KELLER and, consequently, to include an acetylation in our tocopherol determination. We found it to be important to use freshly distilled pyridine and to carry out the acetylation in a N_2 -atmosphere.

We obtained further separation from other reducing non-tocopherols by including in the determination a distillation in a high vacuum. According to EMERSON et al.¹, the tocopherols distill at 10^{-5} mm Hg between 110° and 140° . Our experiments revealed the fact that the unsaponifiable matter of certain natural products, for instance the lipoids of green leaves and hips, contained appreciable amounts of substances which reduce ferric chloride, but do not distill in high vacuum as vitamin E does.

We distilled pure α -tocopherol and mixtures of α -tocopherol and β -carotene for 5 minutes in high vacuum at 140° in the apparatus described below (Fig. 2) and determined the tocopherol-content of the distillate colorimetrically (Table 2).

¹ O. H. EMERSON, G. A. EMERSON, A. MOHAMMED and H. M. EVANS, *J. Biol. Ch.* **122**, 99 (1937).

Table 2.

Product distilled	Per cent recovery
1.9 mg d,l- α -tocopherol	96
1.9 - d,l- α -tocopherol	97
1.9 - d,l- α -tocopherol + 0,17 mg β -carotene..	95
0.95 - d,l- α -tocopherol + 0,68 - β -carotene..	98

Distillation longer than 5 minutes did not yield greater recovery. By distillation of unsaponifiable matter from wheat germ oil the following results were obtained (Table 3). The distillation lasted for 10 minutes.

Table 3.

Product distilled	Per cent recovery
8 mg of unsaponifiable matter of wheat germ oil	94
45 - - - - -	91
7 - - - - -	98
54 - - - - -	94

As "per cent recovery" we denote tocopherol colorimetrically determined in the distillate in relation to tocopherol colorimetrically determined before the distillation. Also here, almost total recovery was obtained if amounts not greater than 10—20 mg were distilled. After acetylation of the unsaponifiable matter of wheat germ oil, practically no reduction power remains.

The unsaponifiable matter of the lipoids of hips and green leaves behaves in quite a different way. Here, a correction of 10—50 % can be obtained by the acetylation method, and a considerably greater correction will be obtained by subtracting that part of the reducing substances which remains in the non-distillable fraction (Table 4).

Table 4.

Product distilled	Colorimetrically determined value calculated as % tocopherol in dried material		
	direct determination	after subtraction for acetylation	in the distillate
4.0 mg of unsaponifiable of hips	0.056	0.048	0.032
4.9 mg — —	0.044	0.033	0.022
6.0 mg — —	0.045	0.034	0.025
5.0 mg of unsaponifiable of spinach	0.261	0.142	0.045
5.2 mg — —	0.272	0.170	0.062
3.0 mg of unsaponifiable of alfalfa	0.192	0.138	0.044
2.0 mg — —	0.144	0.960	0.040

These results suggest the assumption that certain components hitherto unknown must be present in unsaponifiable matter from lipoids of hips and dried leaves, since, if a known amount of d, 1- α -tocopherol is added before the distillation, the total amount of tocopherol can be recovered on the condenser and, furthermore, the total reduction power of distillate + non-distillable is the same as before the distillation.

It might be of interest to investigate the chemical nature of these unknown reducing substances. They will probably be alcohols, since their reducing power is eliminated by acetylation. They have no vitamin E activity when tested on chicks (these studies are reported in Part II of this paper).

If the method is to be applied on animal material, it must be reminded that vitamin A which does not occur in plant material will also reduce ferric chloride. Vita-

min A distills in high vacuum, and the reduction power is diminished by acetylation, however, it may possess part of its reducing properties even after acetylation. Vitamin A is, therefore, not eliminated by the method.

Perhaps, it becomes possible to eliminate vitamin A by means of the antimony trichloride reaction, such as proposed by KARRER and KELLER¹.

Frequently, but a very small acetylation correction is obtained in the distillate; sometimes, this correction may amount to 10 %/o. It is, therefore, advisable in every case to examine whether an acetylation after the distillation is necessary or not.

Procedure for tocopherol-determination in dried plant material.

Production of the unsaponifiable matter. About 5 grams of the material are weighed and ground in a mortar with about equal parts of pulverized quartz and a little CaCO_3 . The mixture is extracted on a glass filter with 33 %/o freshly distilled acetone; this extract is removed. The main extraction is carried out by means of successive portions of pure acetone, until the acetone remains colourless. Then one extraction with 90 %/o acetone and two extractions with ether are carried out. The combined extracts are evaporated in vacuo at about 40° . The residue is transferred to a separatory funnel by means of ether and separated from eventually admixed water. Then the ether is dried with Na_2SO_4 overnight; the Na_2SO_4 is filtered off, washed with ether and the combined extracts are evaporated in vacuo to about 25 cc. The ether solution is then saponified with 4 n methyl alcoholic potassium hydroxide in an N_2 atmosphere in a

¹ loc. cit. p. 7.

separatory funnel for two hours. The mixture is diluted with water, the watery phase is separated and washed with ether. The combined ether phases are washed with water until they are alkali free and are dried over Na_2SO_4 . After filtration and evaporation in vacuo, the residue is soluted in ethyl alcohol plus a little benzene.

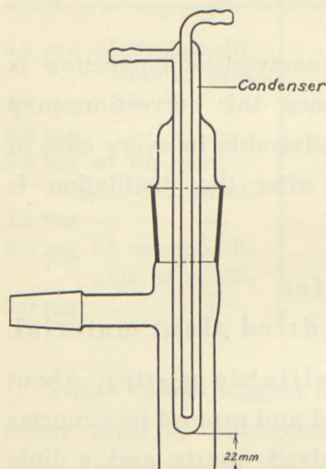


Fig. 2.

Distillation. As much of this solution as contains 5–10 mg of dried material is carefully evaporated on the bottom of the vacuum still (Fig. 2) which is thereafter connected to the high vacuum pump aggregate consisting of a mercury diffusion pump, a condenser immersed in acetone-solid CO_2 and a mechanical fore-vacuum pump. When the pressure has fallen below 10^{-3} mm Hg, measured

by means of a v. Reden manometer, the apparatus is immersed into liquid Wood's metal at 140° so that the surface of the metal is 2.5 cm above the deepest point of the condenser. This temperature is maintained for 25–30 minutes. Then, the metal bath is removed and the apparatus is allowed to cool. The distillate is collected from the condenser by washing with ethanol-benzene.

Acetylation. One part of the distillate is used directly for the colorimetric reaction. The rest is evaporated, soluted in 1 cc acetic anhydride + 5 cc freshly distilled pyridine and the acetylation is carried on for two hours in an N_2 -atmosphere on a boiling water bath with reflux-condenser. The solution is then diluted with petrol ether, and the

petrol ether layer is washed, first with diluted hydrochloric acid and then with water until neutral. The petrol ether is dried with Na_2SO_4 , evaporated and soluted in ethanol-benzene.

The colour reaction. As much of the ethanol-benzene solution as contains about 0.1 mg of tocopherol + 1 cc 0.5 % α, α' -dipyridyl in ethanol + 1 cc 0.2 % FeCl_3 in ethanol is transferred to a 25 cc measuring flask and diluted to the mark with ethanol. The photometer is read after 15 minutes by screen S 50 and 5 cm microcuvettes. The result is calculated by means of the extinction coefficient for pure d, l- α -tocopherol which is 345, as mentioned above.

The tocopherol content of some natural products.

By means of this method, the tocopherol content of a series of natural products was determined (Table 5). It is interesting to see that dried green plants such as spinach, alfalfa, and dried hips contain just as much tocopherol as wheat germ, hitherto looked upon as the most vitamin E-potent source. Lipoid extracts of these products contain much more tocopherol than wheat germ oil. The high

Table 5.

Product	% tocopherol
Wheat germ (after the season)	0.015—0.038
Wheat germ oil (after the season), pressed ...	0.14— 0.25
Wheat germ oil, extracted with petrol ether...	0.45
Wheat germ oil of the same charge pressed ...	0.23
Dried hips (without pips)	0.02— 0.05
Petrol ether extract of dried hips.....about	2.3
Dried spinach	0.05
Dried alfalfa (commercial products)	0.01— 0.045
Apple pips	0.02
Mangold..... less than	0.00001

potency of these products as vitamin E sources has been confirmed in animal experiments (Part II).

Summary.

Various chemical measures for the determination of vitamin E (tocopherols) have been examined. The best results were obtained by means of the α , α' -dipyridyl-ferric chloride method after the product had been purified by saponification and distillation in a high vacuum.

II. Some experiments on estimating the vitamin E activity of natural products by means of alimentary exudative diathesis.

In our previous works on vitamin E, we suggested to use the anti-cephalomalacia activity or the anti-exudation activity of vitamin E for its biological estimation. Such experiments have now been carried out.

Experimental. Chicks weighing about 100 g were kept as usual in individual cages under clean conditions. They were fed the vitamin E-free diet 545 consisting of diet 538 + 5 % cod liver oil.

Diet 538	*) Salt mixture No. 3
Casein, alcohol extracted. 150	Calcium carbonate.... 400
Dried brewer's yeast..... 100	Magnesium carbonate. 20
Sugar 730	Sodium chloride 88
Salt mixture No. 3*) 20	Ferric citrate 32
1000	Di-jodo-tyrosine..... 0.005
	Cupric sulphate, cryst. 3
	Manganous sulphate,
	cryst. 6
	549

Vitamin K was furnished in the form of tablets containing a highly purified product, resp. pure K₁, six times

a week. The substances, whose vitamin E activity had to be tested, resp. pure synthetic d,l- α -tocopherol acetate (kindly put at our disposal by F. Hoffmann-la Roche & Co. A.G., Basel), which was used as standard, were also furnished as tablets six times a week.

Three series of experiments, A, B and C, were carried out.

In series A, the effects of a logarithmic series of doses of dried hips were compared with a logarithmic series of doses of tocopherol. The following doses were used: 0.25 — 0.5 — 1 and 2 g of dried hips and 0.1 — 0.2 — 0.4 — 0.8 — 1.6 and 3.2 mg of d,l- α -tocopherol acetate.

In series B, the effects of varying doses of dried hips were tried again, but smaller doses were used, viz. 60 — 120 and 240 mg. At the same time, the effect of an acetone extract of the dried hips, from which tablets were made by means of lactose and talc, was tested. These tablets were fed in such amounts that the doses corresponded to 60 — 120 and 240 mg of dried hips. From d,l- α -tocopherol acetate doses of 0.1 — 0.2 and 0.4 mg were used.

The experiments in series C were carried out in order to settle the question whether the reducing but non-distilling substances from alfalfa, described in Part I, had any E activity. One group of animals received every day (except Sunday) 1 tablet containing as much of the distillate at 140° and 10⁻³ mm Hg of unsaponifiable matter of alfalfa as, on the basis of the chemical determination, corresponded to 0.4 mg of d,l- α -tocopherol acetate; the second group was fed the corresponding amount of non-distilling product. Further groups got no tablets and 0.1 — 0.2 — 0.4 mg of d,l- α -tocopherol acetate.

The animals were carefully inspected for exudates and encephalomalacia every day. Encephalomalacia was dia-

gnosed by the behaviour of the animals and by the macroscopic findings on opening the brain. After 35—40 days, the surviving animals were killed. Autopsy was made in every case. The results appear from Table 1. In this table, exudations as well as encephalomalacias are indicated by + in the respective columns. As mentioned in our previous papers, the hemorrhagic condition of the fat tissue seen by autopsy is part of the exudative symptom. Where hemorrhage in fat tissue occurred without exudate, the case was considered to be the initial stage of the full symptom. In Table 1, such cases are indicated by (+); by the statistical evaluation (Table 2) they are arbitrarily calculated as $\frac{1}{2}$ full exudation. Table 1 contains, furthermore, the time until the appearance of the exudates as well as the number of days from the beginning of the experiment until the chick died or was killed. Animals which were killed are marked with k, and those which died during the feeding period are marked with d.

The results are summarized in Table 2 which indicates the percentage of animals showing exudates resp. encephalomalacia. Animals which died before the 7th day of the experiment are left out in the calculation.

Table 1.
Series A. Started 28-29/9 1939

Group 1 no addition				Group 2 0.1 mg d,l- α -tocopherol acetate p. day			
Chick No.	Duration of experiment days	Exu-date days	Encephalo-malacia days	Chick No.	Duration of experiment days	Exu-date days	Encephalo-malacia days
5931	27 d	+ 22	+ 14	5821	25 k	(+) ..	0 ..
5932	24 d	+ 20	0 ..	5822	36 k	0 ..	0 ..
5933	4 d	0 ..	0 ..	5823	35 k	0 ..	0 ..
5934	35 k	+ 16	0 ..	5824	13 k	+ 11	0 ..
5935	22 d	+ 8	0 ..	5825	35 k	0 ..	0 ..
5936	12 d	+ 12	0 ..	5826	35 k	0 ..	0 ..
5937	7 d	0 ..	0 ..	5827	35 k	+ 11	0 ..
5938	14 d	+ 12	0 ..	5828	36 k	(+) ..	0 ..
5939	8 d	+ 8	+ 8	5829	33 d	(+) ..	0 ..
5940	8 k	+ 8	+ 8	5830	36 k	+ 11	0 ..
Group 3 0.2 mg d,l- α -tocopherol acetate p. day				Group 4 0.4 mg d,l- α -tocopherol acetate p. day			
5831	37 k	+ 26	0 ..	5841	35 k	0 ..	0 ..
5832	35 k	0 ..	0 ..	5842	35 k	0 ..	0 ..
5833	22 k	(+) ..	0 ..	5843	35 k	0 ..	0 ..
5834	35 k	0 ..	0 ..	5844	36 k	0 ..	0 ..
5835	35 k	0 ..	0 ..	5845	35 k	0 ..	0 ..
5836	36 k	0 ..	0 ..	5846	36 k	0 ..	0 ..
5837	35 k	0 ..	0 ..	5847	36 k	0 ..	0 ..
5838	37 k	+ 20	0 ..	5848	36 k	0 ..	0 ..
5839	36 k	0 ..	0 ..	5849	37 k	0 ..	0 ..
5840	36 k	+ 19	0 ..	5850	37 k	0 ..	0 ..
Group 5 0.8 mg d,l- α -tocopherol acetate p. day				Group 6 1.6 mg d,l- α -tocopherol acetate p. day			
5851	35 k	0 ..	0 ..	5861	35 k	0 ..	0 ..
5852	35 k	0 ..	0 ..	5862	35 k	0 ..	0 ..
5853	36 k	0 ..	0 ..	5863	35 k	0 ..	0 ..
5854	35 k	0 ..	0 ..	5864	36 k	0 ..	0 ..
5855	35 k	0 ..	0 ..	5865	35 k	0 ..	0 ..
5856	12 d	0 ..	0 ..	5866	36 k	0 ..	0 ..
5857	36 k	0 ..	0 ..	5867	36 k	0 ..	0 ..
5858	36 k	0 ..	0 ..	5868	36 k	0 ..	0 ..
5859	36 k	0 ..	0 ..	5869	35 d	0 ..	0 ..
5860	37 k	0 ..	0 ..	5870	6 d	0 ..	0 ..

Table 1 (continued).

Group 7 3.2 mg d,l- α -tocopherol acetate p. day				Group 8 0.25 g dried hips p. day			
Chick No.	Duration of experiment days	Exu-date days	Ence-phalo-mala-cia days	Chick No.	Duration of experiment days	Exu-date days	Ence-phalo-mala-cia days
5871	35 k	0 ..	0 ..	5881	35 k	0 ..	0 ..
5872	35 k	0 ..	0 ..	5882	35 k	(+) ..	0 ..
5873	35 k	0 ..	0 ..	5883	35 k	0 ..	0 ..
5874	36 k	0 ..	0 ..	5884	35 k	0 ..	0 ..
5875	27 d	0 ..	0 ..	5885	15 d	(+) ..	0 ..
5876	36 k	0 ..	0 ..	5886	36 k	0 ..	0 ..
5877	36 k	0 ..	0 ..	5887	36 k	0 ..	0 ..
5878	36 k	0 ..	0 ..	5888	37 k	(+) ..	0 ..
5879	37 k	0 ..	0 ..	5889	37 k	0 ..	0 ..
5880	36 k	0 ..	0 ..	5890	37 k	0 ..	0 ..
Group 9 0.5 g dried hips p. day				Group 10 1 g dried hips p. day			
5891	35 k	0 ..	0 ..	5901	35 k	0 ..	0 ..
5892	35 k	0 ..	0 ..	5902	36 k	0 ..	0 ..
5893	35 k	0 ..	0 ..	5903	35 k	0 ..	0 ..
5894	35 k	(+) ..	0 ..	5904	35 k	0 ..	0 ..
5895	36 k	0 ..	0 ..	5905	36 k	0 ..	0 ..
5896	36 k	0 ..	0 ..	5906	35 k	0 ..	0 ..
5897	37 k	0 ..	0 ..	5907	15 d	(+) ..	0 ..
5898	37 k	0 ..	0 ..	5908	2 d	0 ..	0 ..
5899	37 k	0 ..	0 ..	5909	36 k	0 ..	0 ..
5900	11 d	0 ..	0 ..	5910	36 k	0 ..	0 ..
Group 11 2 g dried hips p. day							
5912	35 k	0 ..	0 ..				
5913	35 k	0 ..	0 ..				
5914	35 k	0 ..	0 ..				
5915	3 d	0 ..	0 ..				
5916	35 k	0 ..	0 ..				
5917	36 k	0 ..	0 ..				
5919	20 d	0 ..	0 ..				
5920	36 k	0 ..	0 ..				
5922	37 k	0 ..	0 ..				
5925	37 k	0 ..	0 ..				
5926	37 k	0 ..	0 ..				
5927	37 k	0 ..	0 ..				

Table 1 (continued).
Series B. Started 20/11-1939.

Group 1 no addition				Group 2 0.1 mg d,l- α -tocopherol acetate p. day			
Chick No.	Dura- tion of experi- ment days	Exu- date days	Ence- phalo- mala- cia days	Chick No.	Dura- tion of experi- ment days	Exu- date days	Ence- phalo- mala- cia days
5961	28 k	+ 28	+ 16	5971	40 k	0 ..	0 ..
5962	24 k	+ 16	0 ..	5972	40 k	+ 40	0 ..
5963	14 d	+ 14	0 ..	5973	40 k	0 ..	0 ..
5964	16 k	0 ..	0 ..	5974	40 k	+ 29	+ 35
5965	40 k	+ 40	0 ..	5975	43 k	0 ..	0 ..
5966	16 d	+ 14	0 ..	5976	43 k	0 ..	0 ..
5967	25 k	+ 14	+ 19	5977	43 k	0 ..	0 ..
5968	12 d	(+) ..	0 ..	5978	43 k	0 ..	0 ..
5969	12 d	(+) ..	0 ..	5979	43 k	+ 25	0 ..
5970	40 k	0 ..	0 ..	5980	43 k	+ 10	0 ..
Group 3 0.2 mg d,l- α -tocopherol acetate p. day				Group 4 0.4 mg d,l- α -tocopherol acetate p. day			
5981	40 k	0 ..	0 ..	5991	40 k	0 ..	0 ..
5982	40 k	+ 29	0 ..	5992	40 k	0 ..	0 ..
5983	40 k	0 ..	0 ..	5993	40 k	0 ..	0 ..
5984	43 k	0 ..	0 ..	5994	38 k	0 ..	0 ..
5985	43 k	(+) ..	0 ..	5995	38 k	0 ..	0 ..
5986	43 k	0 ..	0 ..	5996	40 k	0 ..	0 ..
5987	43 k	(+) ..	0 ..	5997	38 k	0 ..	0 ..
5988	38 k	+ 38	0 ..	5998	38 k	0 ..	0 ..
5989	43 k	0 ..	0 ..	5999	43 k	0 ..	0 ..
5990	43 k	0 ..	0 ..	7000	42 k	0 ..	0 ..
Group 5 1 tablet of dried hips (60 mg) p. day				Group 6 2 tablets of dried hips (60 mg each) p. day			
7001	40 k	0 ..	0 ..	7011	40 k	+ 30	0 ..
7002	40 k	0 ..	+ 25	7012	10 d	+ 8	0 ..
7003	40 k	0 ..	0 ..	7013	2 d	0 ..	0 ..
7004	38 k	+ 19	0 ..	7014	40 k	0 ..	0 ..
7005	20 d	0 ..	+ 19	7015	40 k	0 ..	0 ..
7006	42 k	0 ..	0 ..	7016	41 d	+ 39	0 ..
7007	38 k	+ 31	0 ..	7017	42 k	(+) ..	0 ..
7008	42 k	0 ..	0 ..	7018	10 k	+ 9	0 ..
7009	42 k	0 ..	0 ..	7019	42 k	0 ..	0 ..
7010	42 k	+ 22	0 ..	7020	42 k	0 ..	0 ..

Table 1 (continued).

Group 7 4 tablets of dried hips (60 mg each) p. day				Group 8 1 tablet (60 mg) of extract of dried hips p. day			
Chick No.	Dura- tion of experiment days	Exu- date days	Ence- phalo- mala- cia days	Chick No.	Dura- tion of experiment days	Exu- date days	Ence- phalo- mala- cia days
7021	40 k	(+) ..	0 ..	7031	22 d	0 ..	+ 10
7022	24 k	+ 22	0 ..	7032	13 d	+ 13	0 ..
7023	40 k	0 ..	0 ..	7033	40 k	0 ..	0 ..
7024	40 k	+ 30	0 ..	7034	40 k	0 ..	0 ..
7025	42 k	0 ..	0 ..	7035	37 k	+ 36	0 ..
7026	42 k	+ 42	0 ..	7036	40 k	0 ..	+ 23
7027	42 k	0 ..	0 ..	7037	42 k	0 ..	0 ..
7028	28 k	+ 23	0 ..	7038	42 k	+ 22	0 ..
7029	42 k	+ 39	0 ..	7039	16 k	+ 9	+ 10
7030	42 k	0 ..	0 ..	7040	42 k	+ 24	0 ..
Group 9 2 tablets (60 mg each) of extract of dried hips p. day				Group 10 4 tablets (60 mg each) of extract of dried hips p. day			
7041	10 k	+ 8	0 ..	7051	40 k	0 ..	0 ..
7042	40 k	0 ..	0 ..	7052	7 d	0 ..	0 ..
7043	40 k	0 ..	0 ..	7053	38 k	+ 38	0 ..
7044	40 k	0 ..	0 ..	7054	40 k	0 ..	0 ..
7045	42 k	+ 37	0 ..	7055	40 k	(+) ..	0 ..
7046	26 k	+ 15	0 ..	7056	42 k	(+) ..	0 ..
7047	42 k	0 ..	0 ..	7057	42 k	0 ..	0 ..
7048	42 k	0 ..	0 ..	7058	21 d	0 ..	+ 19
7049	42 k	0 ..	+ 18	7059	42 k	(+) ..	0 ..
7050	42 k	+ 23	0 ..	7060	42 k	0 ..	0 ..

Series C. Started 16/5-1940.

Group 1 no addition				Group 2 0.1 mg d,l- α -tocopherol acetate p. day			
9080	39 d	+ 31	0 ..	9090	39 k	0 ..	0 ..
9081	34 d	+ 27	0 ..	9091	39 k	0 ..	0 ..
9082	39 k	0 ..	0 ..	9092	39 k	0 ..	0 ..
9083	39 k	0 ..	0 ..	9093	35 d	0 ..	0 ..
9084	39 k	+ 39	0 ..	9094	39 k	0 ..	0 ..
9085	39 k	+ 27	+ 36	9095	39 k	0 ..	0 ..
9086	36 k	+ 29	0 ..	9096	40 k	+ 29	0 ..
9087	30 d	+ 16	0 ..	9097	40 k	0 ..	0 ..
9088	40 k	0 ..	0 ..	9098	40 k	0 ..	0 ..
9089	40 k	0 ..	0 ..	9099	40 k	0 ..	0 ..

Table 1 (continued).

Group 3 0.2 mg d,l- α -tocopherol acetate p. day				Group 4 0.4 mg d,l- α -tocopherol acetate p. day			
Chick No.	Duration of experiment days	Exu-date days	Encephalomalacia days	Chick No.	Duration of experiment days	Exu-date days	Encephalomalacia days
9100	39 k	0 ..	0 ..	9110	39 k	0 ..	0 ..
9101	39 k	0 ..	0 ..	9111	39 k	0 ..	0 ..
9102	39 k	0 ..	0 ..	9112	39 k	0 ..	0 ..
9103	39 k	0 ..	0 ..	9113	29 d	0 ..	0 ..
9104	39 k	0 ..	0 ..	9114	39 k	0 ..	0 ..
9105	39 k	0 ..	0 ..	9115	39 k	0 ..	0 ..
9106	40 k	0 ..	0 ..	9116	40 k	0 ..	0 ..
9107	40 k	0 ..	0 ..	9117	40 k	0 ..	0 ..
9108	40 k	0 ..	0 ..	9118	40 k	0 ..	0 ..
9109	40 k	0 ..	0 ..	9119	40 k	0 ..	0 ..
Group 5 distillate from alfalfa				Group 6 undistillable from alfalfa			
7441	39 k	0 ..	0 ..	7453	39 k	+ 27	0 ..
7442	39 k	0 ..	0 ..	7454	39 k	+ 32	0 ..
7443	38 d	0 ..	0 ..	7455	39 k	+ 25	0 ..
7444	39 k	0 ..	0 ..	7456	39 k	+ 28	0 ..
7445	30 d	0 ..	0 ..	7457	39 k	0 ..	0 ..
7446	39 k	0 ..	0 ..	7458	39 k	0 ..	0 ..
7447	39 k	0 ..	0 ..	7459	40 k	+ 25	0 ..
7448	40 k	0 ..	0 ..	7460	40 k	+ 30	0 ..
7449	40 k	0 ..	0 ..	7461	40 k	0 ..	0 ..
7450	40 k	0 ..	0 ..	7462	38 d	+ 34	0 ..
7451	40 k	0 ..	0 ..	7463	40 k	+ 40	0 ..
7452	40 k	0 ..	0 ..	7464	40 k	+ 21	+ 11

Table 2.
Series A.

Group No.	Addition	% of animals with		Total percentage of animals with E avitaminous symptoms
		exudative diathesis	encephalomalacia	
1	no	100	38	100
2	0.1 mg d,l- α -tocopherol acetate	45	0	45
3	0.2 - — — —	35	0	35
4	0.4 - — — —	0	0	0
5	0.8 - — — —	0	0	0
6	1.6 - — — —	0	0	0
7	3.2 - — — —	0	0	0
8	250 - dried hips	15	0	15
9	500 - — —	5	0	5
10	1000 - — —	5	0	5
11	2000 - — —	0	0	0

Series B.

1	no	70	20	70
2	0.1 mg d,l- α -tocopherol acetate	40	10	40
3	0.2 - — — —	30	0	30
4	0.4 - — — —	0	0	0
5	60 - dried hips	30	20	50
6	120 - — —	45	0	45
7	240 - — —	55	0	55
8	60 - extract of dried hips..	50	30	70
9	120 - — — — ..	40	10	50
10	240 - — — — ..	28	11	39

Series C.

1	no	60	10	60
2	0.1 mg d,l- α -tocopherol acetate	10	0	10
3	0.2 - — — —	0	0	0
4	0.4 - — — —	0	0	0
5	.. distillate from alfalfa	0	0	0
6	.. undistillable from alfalfa..	75	8	75

Results and discussion. The results show that it is possible to use the chick for a preventive method of standardization of vitamin E by means of the vitamin's anti-exudative, resp. anti-encephalomalacia activity. The results indicate that the exudative symptom gives the most graduated relation between the dosage of the substance and the response; the evaluation of the experiments can therefore be carried out by means of the exudative symptom alone. The encephalomalacia symptom alone may be used under other circumstances, when diets richer in fat are given. In the present case, however, we preferred to calculate the total number of animals with symptoms of E avitaminosis.

When the basal diet without addition of vitamin E is fed (see, for instance, Table 3), the symptoms occur in 60—100 % (an average of 80 %) of the animals. A comparison between the activity of two different sources of vitamin E can therefore be carried out in such a way that both substances are fed at the same time in different levels to different groups of chicks, and the doses are calculated which give symptoms by 40 % of the animals. As is the case in similar standardizations, it is necessary to test, simultaneously with every standardization, the reference standard, d,l- α -tocopherol acetate, since the response to a given dose of the vitamin fluctuates, even if animals of the same stock and on the same diet are used. Table 3 shows the reproducibility of the chick's response to the same series of doses of the reference substance. These series are collected from the experiments in this and in the following parts of the paper.

The experiments show that the dosis of the reference substance which gives symptoms by 40 % of the animals is about 0.1 mg per day.

Table 3.

Series No.	Date of experiment	Duration of experiment days	mg of d,l- α -tocopherol acetate p. day			
			0	0.1	0.2	0.4
			percentage of chicks with symptoms			
II A	28/9-39	about 35	100	45	35	0
II B	20/11-39	— 40	70	40	30	0
II C	16/5-40	— 39	60	60	5	0
III B	24/4-40	— 35	90	35	25	0

In series B, we did not feed a dose of dried hips and extract of dried hips sufficient to yield 100 % protection; furthermore, the results for dried hips were unusually irregular, whereas extract of dried hips gave more regular results. If we try nevertheless to find the dosis showing symptoms by 40 % of the animals, the results of series B lead to an amount of about 240 mg of extract of dried hips per day, and B conferred with A gives about 240 mg of dried hips per day, in the case of d,l- α -tocopherol acetate the amount is 0.1 mg. This result is in rather good agreement with the chemical determinations described in Part I, which showed a tocopherol content of 0.124 mg in 240 mg hips and of 0.092 mg in the corresponding amount of extract, calculated as acetate; the values are of the same order of magnitude as those found in rat fertility tests in earlier experiments with hips. Even if the result is no exact standardization, it shows nevertheless that this method enables us to carry out vitamin E standardizations. If exact standardizations are to be carried out, the number of days until the symptoms occur should be taken into consideration. If the amount of vitamin E ingested is increased, the number of animals showing symptoms decreases and, besides, also the number of days till the appearance of the symptoms increases.

Summary.

The principles of a method of the biological standardization of vitamin E by means of its anti-exudative and anti-encephalomalacia activity are described. By means of this method it was tried to determine the vitamin E activity of hips.

III. Comparison between the anti-exudative activity of α - und β -tocopherol.

The standardization method of Part II was applied in order to investigate whether the relation between the potency of α - and β -tocopherol in preventing exudative diathesis is the same as the relation between the potencies found in rat fertility tests.

Experimental. The experimental conditions with respect to keeping of the animals, diet, inspection for symptoms, autopsy etc. were the same as described in Part II. The results are summarized in tables with the same indications + and (+) for exudation symptoms as in Part II. The animals were inspected for encephalomalacia too, but no case showing this symptom could be found.

Two series of experiments, A and B, were carried out.

In series A, the effects of equal doses of α - and β -tocopherol acetates were compared. The tocopherol acetates were given in the form of tablets every day except Sunday. Each tablet contained 0.1 mg of the tocopherol acetate. The dose was 0.4 mg per animal per day until the weight had increased to 150 g. Thereafter, the dose increased by 50 % for every 50 g gain in weight over 100 g. The synthetic d,l- α -tocopherol acetate was supplied from F. Hoffmann-La Roche & Co. A.G., Basle. Synthetic β -tocopherol acetate was kindly put at our disposal by Prof. P. Karrer, Zürich.

In series B, the effects of varying doses of α - and β -tocopherol acetates were tested. The following doses were used: 0.05 — 0.1 — 0.2 — 0.4 mg of d,l- α - and 0.4 — 0.8 — 1.6 mg of d,l- β -tocopherol acetate per animal per day. The doses were kept constant through the whole period of experi-

Table 1.
Series A. Started 5/7-1939.

Group 1 No addition of tocopherol acetate			Group 2 Addition of d,l- α - tocopherol acetate			Group 3 Addition of d,l- β - tocopherol acetate		
Chick No.	Dura- tion of experi- ment days	Exu- date days	Chick No.	Dura- tion of experi- ment days	Exu- date days	Chick No.	Dura- tion of experi- ment days	Exu- date days
5563	6 d	0 ..	5573	40 k	0 ..	5583	19 k	+ 15
5564	5 d	0 ..	5574	40 k	0 ..	5584	17 d	0 ..
5565	14 k	+ 13	5575	40 k	0 ..	5585	15 k	+ 14
5566	29 d	0 ..	5576	40 k	+ 7	5586	28 d	+ 27
5567	35 k	+ 31	5577	40 k	0 ..	5587	5 d	0 ..
5568	20 k	+ 19	5578	40 k	0 ..	5588	11 d	0 ..
5569	20 k	+ 17	5579	38 k	0 ..	5589	27 k	+ 24
5570	15 k	+ 15	5580	9 d	0 ..	5590	35 k	+ 35
5571	15 k	+ 14	5581	40 k	0 ..	5591	19 d	0 ..
5572	9 k	+ 7	5582	40 k	0 ..	5592	20 k	+ 18

Series B. Started 24/4-1940.

Group 1 No addition of tocopherol acetate			Group 2 Addition of 0.05 mg d,l- α -tocopherol acetate			Group 3 Addition of 0.1 mg d,l- α -tocopherol acetate		
7401	34 k	+ 26	7391	34 k	0 ..	7381	34 k	0 ..
7402	34 k	+ 27	7392	34 k	+ 34	7382	34 k	0 ..
7403	33 d	+ 12	7393	34 k	(+) ..	7383	34 k	+ 22
7404	34 k	+ 24	7394	34 k	+ 26	7384	34 k	0 ..
7405	16 d	+ 16	7395	34 k	+ 15	7385	34 k	(+) ..
7406	34 k	+ 22	7396	34 k	0 ..	7386	34 k	0 ..
7407	35 k	+ 21	7397	35 k	+ 35	7387	35 k	+ 21
7408	35 k	+ 28	7398	35 k	+ 26	7388	29 d	+ 23
7409	24 d	+ 21	7399	35 k	+ 27	7389	35 k	0 ..
7410	35 k	0 ..	7400	32 d	(+) ..	7390	35 k	0 ..

Table 1 (continued).

Group 4 Addition of 0.2 mg d,l- α -tocopherol acetate			Group 5 Addition of 0.4 mg d,l- α -tocopherol acetate			Group 6 Addition of 0.4 mg d,l- β -tocopherol acetate		
Chick No.	Duration of experi- ment days	Exu- date days	Chick No.	Duration of experi- ment days	Exu- date days	Chick No.	Duration of experi- ment days	Exu- date days
7371	34 k	0 ..	7361	34 k	0 ..	7431	34 k	+ 34
7372	34 k	0 ..	7362	34 k	0 ..	7432	34 k	+ 34
7373	34 k	(+)	7363	34 k	0 ..	7433	34 k	+ 22
7374	34 k	0 ..	7364	34 k	0 ..	7434	34 k	+ 33
7375	34 k	0 ..	7365	34 k	0 ..	7435	34 k	0 ..
7376	34 k	0 ..	7366	22 d	0 ..	7436	34 k	+ 34
7377	35 k	+ 35	7367	35 k	0 ..	7437	35 k	+ 33
7378	35 k	0 ..	7368	35 k	0 ..	7438	35 k	+ 13
7379	35 k	0 ..	7369	35 k	0 ..	7439	35 k	+ 28
7380	35 k	+ 35	7390	35 k	0 ..	7440	35 k	+ 33
Group 7 Addition of 0.8 mg d,l- β -tocopherol acetate			Group 8 Addition of 1.6 mg d,l- β -tocopherol acetate					
7421	16 d	0 ..	7411	34 k	+ 12			
7422	34 k	+ 26	7412	16 d	0 ..			
7423	34 k	+ 20	7413	34 k	+ 26			
7424	34 k	+ 22	7414	34 k	+ 21			
7425	34 k	0 ..	7415	34 k	0 ..			
7426	34 k	+ 33	7416	34 k	+ 23			
7427	35 k	+ 21	7417	35 k	+ 33			
7428	25 d	+ 12	7418	35 k	+ 21			
7429	35 k	+ 20	7419	35 k	0 ..			
7430	35 k	+ 33	7420	32 d	+ 17			

ment. The β -tocopherol acetate applied in this experiment was prepared in our laboratory following the method of KARRER and co-workers¹. The phytol used in the synthesis was kindly supplied by F. Hoffmann-la Roche & Co. A.G. and Chemische Fabrik vormals Sandoz, Basle. The β -tocopherol was acetylated and again purified by chromatographic

¹ P. KARRER, H. KOENIG, B. H. RINGIER and H. SALOMON, *Helv. chim. acta* **22**, 1139 (1939).

Table 2.
Series A.

Addition	% of animals with exudate
no	88
d,l- α -tocopherol acetate	10
d,l- β -tocopherol acetate	67

Series B.

no	90
0.05 mg d,l- α -tocopherol acetate	70
0.1 mg —	35
0.2 mg —	25
0.4 mg —	0
0.4 mg d,l- β -tocopherol acetate	90
0.8 mg —	80
1.6 mg —	70

Table 3.

Addition	Weight g					
	24/4	1/5	8/5	15/5	22/5	28/5
no	96	122	147	191	229	229
0.05 mg d,l- α -tocopherol acetate	91	110	133	172	222	241
0.1 mg —	91	112	132	170	191	205
0.2 mg —	89	111	126	172	210	234
0.4 mg —	89	106	128	163	193	210
0.4 mg d,l- β -tocopherol acetate	100	120	132	177	211	241
0.8 mg —	91	115	136	181	200	215
1.6 mg —	89	118	144	202	231	251

absorption on aluminium oxide. With respect to its absorption spectrum it agreed with the spectrum of neo-tocopherol as found by KARRER¹. Furthermore, its equivalent weight, determined by reduction of FeCl₃-dipyridyl after EMMERIE and ENGEL, agreed with its formula.

The animals were weighed once a week. In series A, the animals were killed as soon as they showed exudates;

¹ P. KARRER, H. SALOMON and H. FRITZSCHE, *Helv. chim. acta* **21**, 309 (1938).

in series B, however, they were kept alive until the 34th day. A summary of the weights of the groups of this series is given in Table 3.

Results and discussion. Table 2, series A, clearly shows that there is a great difference in the effectiveness of α - and β -tocopherol acetates in preventing the exudative diathesis in chicks.

In series B, it was attempted to evaluate the relative activity of the two compounds by testing the effects of a series of doses of each of them. From Table 2, series B, it appears, however, that the potency of β -tocopherol acetate is so small that even a dose which was four times as great as the completely protecting dose of the α -compound did not exert any protective effect at all.

Since a dose of β -tocopherol acetate sufficient to protect against the symptom has not been tested, it is impossible from the results obtained in these experiments to determine the relative anti-exudative potency of the components. The results, however, suggest that the potency of the β -compound is less than 5 % of that of the α -compound.

Previous experiments carried out in several laboratories have shown that α -tocopherol has a potency in preventing rat sterility of 3—4 times that of β -tocopherol. We compared the potencies of our preparations used in series B by means of this test too. This experiment was kindly carried out by cand. pharm. Kjerbye-Nielsen, A/S Medicinalco, following the method used in this laboratory. 0.7 mg of d,l- α -tocopherol acetate gave 65 % protection; 4 mg of d,l- β -tocopherol acetate gave 100 % protection, 2 mg and 1 mg gave 0 % protection, which is in accordance with the earlier experiments.

In the chick experiment, however, it was found that 0.05 mg of d,l- α -tocopherol acetate was more potent than 1.6 mg of d,l- β -tocopherol acetate, i. e. that α -tocopherol is more than 30 times as potent as β -tocopherol. It is, therefore, possible that the relative potency of the E vitamins in preventing the most characteristic symptom of E avitaminosis in chicks is not the same as the potency of the same compounds in preventing rat sterility.

If this statement holds, it would be an analogy of the properties of the D vitamins, where the antirachitic potency of the vitamins D₂ and D₃ is not the same when tested on rats or on chicks.

It would be of great interest to reinvestigate this question by using a greater number of chicks and higher doses of β -tocopherol than applied in this experiment.

It would also be of interest to investigate the anti-encephalomalacia-activity of β -tocopherol acetate in relation to that of the α -tocopherol acetate. This would afford a diet much richer in fat than the diet used in this experiment in which the presence of 5 % cod liver oil was not sufficient to develop cases of encephalomalacia.

From Table 3 it appears that d,l- α -tocopherol acetate has no influence on the growth of chicks, when this diet is used. In earlier experiments, however, when other diets were used, we have seen a significant influence of tocopherol upon the growth of the chicks. It would be of interest to examine this question further and especially to investigate the influence of vitamin E on the growth of chicks being fed with a diet rich in fat.

Summary.

Whilst d,l- α -tocopherol acetate is 3—4 times as active as d,l- β -tocopherol acetate in preventing rat sterility, it was found to be at least 30 times as active as the β -compound in preventing exudative diathesis in chicks.

IV. The influence of fat on the development of encephalomalacia and exudative diathesis in chicks.

In accordance with the experience of PAPPENHEIMER and GOETTSCH and their co-workers¹, we observed that the encephalomalacia occurs in a greater percentage of the experimental animals when the diet contains much fat, a fact which is also in agreement with the still earlier experience that the most commonly known symptom of E avitaminosis, viz. the sterility, is particularly easy to develop by means of a fat rich diet.

As regards the exudative diathesis, our earlier experiments did not reveal any large difference in the percentage of animals afflicted with this symptom, dependent on whether the diet contained extra fat or not.

Recently, we had an opportunity of investigating the influence of fat on both symptoms on a larger scale. The results of these experiments are reported in the following.

Experimental. Experimental conditions with respect to housing of the animals, inspection for symptoms etc. were the same as described in Part II. As basal diet we used diet 538 (cfr. Part II). Other diets were prepared by addition of lard resp. cod liver oil to diet 538.

Vitamins A and D were given in the form of a mixture

¹ A. M. PAPPENHEIMER, MARIANNE GOETTSCH and E. JUNGHERR, Storrs Agricultural Experiment Station Bulletin **229** (Jan. 1939).

Table 1.

Series A Started 8/1-1940				Series B Started 9/3-1940			
Chick No.	Duration of experiment days	Exu-date days	Ence-phalo-mala-cia days	Chick No.	Duration of experiment days	Exu-date days	Ence-phalo-mala-cia days
Group 1: Diet 538 without addition of lard							
7201	49 k	0 ..	+ 47	7241	49 k	+ 31	0 ..
7202	49 k	+ 39	0 ..	7242	49 k	0 ..	0 ..
7203	49 k	(+) ..	0 ..	7243	46 k	0 ..	0 ..
7204	50 k	0 ..	0 ..	7244	49 k	0 ..	0 ..
7205	50 k	+ 35	0 ..	7245	49 k	0 ..	0 ..
7206	50 k	+ 50	0 ..	7246	51 k	0 ..	0 ..
7207	4 d	0 ..	0 ..	7247	51 k	(+) ..	0 ..
7208	49 k	0 ..	0 ..	7248	51 k	0 ..	0 ..
7209	49 k	+ 49	0 ..	7249	51 k	0 ..	0 ..
7210	49 k	0 ..	0 ..	7250	52 k	0 ..	+ 30
7211	49 k	0 ..	0 ..	7251	2 d	0 ..	0 ..
7212	36 d	0 ..	0 ..	7252	52 k	0 ..	0 ..
Group 2: Diet 538 + 1 ⁰ % of lard.							
7143	49 k	+ 49	0 ..	7253	49 k	0 ..	0 ..
7144	49 k	0 ..	0 ..	7254	49 k	0 ..	0 ..
7145	49 k	0 ..	0 ..	7255	49 k	0 ..	0 ..
7146	49 k	0 ..	0 ..	7256	49 k	0 ..	0 ..
7147	50 k	+ 44	0 ..	7257	49 k	0 ..	0 ..
7148	49 k	0 ..	0 ..	7258	51 k	0 ..	0 ..
7149	50 k	(+) ..	0 ..	7259	41 d	+ 16	0 ..
7150	50 k	(+) ..	0 ..	7260	51 k	+ 31	0 ..
7151	49 k	0 ..	+ 49	7261	52 k	0 ..	0 ..
7152	49 k	0 ..	0 ..	7262	52 k	0 ..	0 ..
7153	49 k	0 ..	0 ..	7263	52 k	0 ..	0 ..
7154	49 k	0 ..	0 ..	7264	22 d	0 ..	0 ..
Group 3: Diet 538 + 3 ⁰ % of lard.							
7155	49 k	0 ..	0 ..	7265	49 k	0 ..	0 ..
7156	49 k	0 ..	0 ..	7266	49 k	0 ..	0 ..
7157	44 d	+ 44	0 ..	7267	49 k	0 ..	0 ..
7158	49 k	0 ..	0 ..	7268	49 k	+ 37	0 ..
7159	7 d	0 ..	0 ..	7269	32 d	0 ..	0 ..
7160	50 k	0 ..	0 ..	7270	51 k	0 ..	0 ..

Table 1 (continued).

Series A Started 8/1-1940				Series B Started 9/3-1940			
Chick No.	Duration of experiment days	Exu-date days	Encephalomalacia days	Chick No.	Duration of experiment days	Exu-date days	Encephalomalacia days

Group 3 (continued).

7161	50 k	0 ..	0 ..	7271	51 k	+ 49	0 ..
7162	49 k	0 ..	0 ..	7272	51 k	+ 48	0 ..
7163	49 k	0 ..	0 ..	7273	51 k	+ 48	0 ..
7164	49 k	0 ..	0 ..	7274	52 k	(+) ..	0 ..
7165	49 k	0 ..	+ 49	7275	52 k	0 ..	0 ..
7166	49 k	+ 39	0 ..	7276	52 k	0 ..	0 ..

Group 4: Diet 538 + 9% of lard.

7167	49 k	0 ..	0 ..	7277	49 k	0 ..	0 ..
7168	49 k	0 ..	0 ..	7278	49 k	+ 42	0 ..
7169	37 d	0 ..	0 ..	7279	49 k	+ 49	0 ..
7170	49 k	0 ..	0 ..	7280	49 k	0 ..	0 ..
7171	45 d	+ 36	0 ..	7281	51 k	0 ..	+ 49
7172	43 d	+ 15	0 ..	7282	51 k	+ 45	0 ..
7173	50 k	(+) ..	0 ..	7283	51 k	0 ..	0 ..
7174	49 k	+ 41	0 ..	7284	51 k	0 ..	0 ..
7175	49 k	+ 44	0 ..	7285	52 k	+ 46	0 ..
7176	49 k	0 ..	0 ..	7286	52 k	0 ..	0 ..
7177	49 k	0 ..	0 ..	7287	52 k	+ 52	0 ..
7178	49 k	0 ..	0 ..	7288	52 k	+ 52	0 ..

Group 5: Diet 538 + 27% of lard.

7179	28 d	+ 22	0 ..	7289	49 k	+ 34	+ 44
7180	47 d	+ 40	0 ..	7290	49 k	0 ..	+ 45
7181	26 d	(+) ..	+ 21	7291	49 k	+ 21	0 ..
7182	49 k	0 ..	0 ..	7292	49 k	+ 26	+ 45
7183	49 k	+ 41	+ 46	7293	23 d	+ 11	+ 22
7184	49 k	+ 43	0 ..	7294	51 k	+ 47	+ 22
7185	39 d	+ 37	0 ..	7295	51 k	0 ..	+ 22
7186	43 k	(+) ..	0 ..	7296	51 k	0 ..	0 ..
7187	48 k	(+) ..	+ 30	7297	51 k	0 ..	+ 42
7188	48 k	0 ..	0 ..	7298	52 k	+ 52	0 ..
7189	43 d	+ 32	+ 20	7299	52 k	0 ..	0 ..
7190	24 d	0 ..	+ 17	7300	52 k	+ 46	0 ..

Table 2.

% of lard	Series A			Series B		
	% of animals			% of animals		
	with exudation	with encephalomalacia	surviving the 49th day	with exudation	with encephalomalacia	surviving the 49th day
0	41	9	91	14	9	100
1	25	8	100	17	0	83
3	8	9	100	38	0	92
9	38	0	83	50	8	100
27	63	42	50	58	58	92

Table 3.
Series A.

% of lard	Weight g							
	8/1	13/1	20/1	27/1	3/2	10/2	17/2	24/2
0	95	106	135	170	208	236	264	293
1	100	112	142	176	219	259	291	326
3	101	109	139	179	207	242	264	284
9	101	112	136	163	182	200	233	228
27	96	102	117	139	145	165	180	189
1 mg d,1- α -tocopherol acetate.	95	104	138	177	207	244	265	301

Series B.

% of lard	Weight g							
	9/3	16/3	23/3	30/3	6/4	13/4	20/4	27/4
0	119	117	151	193	247	274	295	340
1	112	132	164	207	269	294	307	357
3	114	136	176	242	300	336	369	417
9	118	134	163	197	247	281	323	356
27	110	118	134	151	160	183	193	205

Table 4.

Group 1 Diet 538 + 5% of lard				Group 2 Diet 538 + 5% of cod liver oil			
Started 16/5-1940							
Chick No.	Duration of experiment days	Exu-date days	Encephalomalacia days	Chick No.	Duration of experiment days	Exu-date days	Encephalomalacia days
9070	39 k	0 ..	0 ..	9080	39 d	+ 31	0 ..
9071	39 k	0 ..	0 ..	9081	34 d	+ 27	0 ..
9072	39 k	0 ..	0 ..	9082	39 k	0 ..	0 ..
9073	39 k	0 ..	+ 14	9083	39 k	0 ..	0 ..
9074	34 d	+ 30	0 ..	9084	39 k	+ 39	0 ..
9075	40 k	+ 36	0 ..	9085	39 k	+ 27	+ 33
9076	35 d	+ 30	0 ..	9086	36 k	+ 29	0 ..
9077	40 k	+ 33	0 ..	9087	30 d	+ 16	0 ..
9078	40 k	+ 24	0 ..	9088	40 k	0 ..	0 ..
9079	40 k	0 ..	0 ..	9089	40 k	0 ..	0 ..

Summary

	% of animals	
	with exudative diathesis	with encephalomalacia
Diet 538 with 5% of lard	50	10
Diet 538 with 5% of cod liver oil..	60	10

of 1 part of Vogan + 2 parts of Vigantol (Bayer); 0.05 cc three times a week.

Vitamin K was supplied as tablets containing 0.005 mg of pure K_1 per tablet. One tablet was given 6 days of the week. For every 50 g gain in weight, the dose was increased by 50 %.

Two series of experiments, A and B, were carried out with the same diets and under the same experimental conditions; they were started with an interval of 2 months.

The results are summarized in Tables 1 and 2 which are arranged as in Part II making use of the same indications.

As already mentioned, the animals were weighed once a week. Even if the ratio between protein and total calories was not kept constant from one diet to another, it may, perhaps, be of interest to give a summary of the weights. This is done in Table 3.

In series A, the group without lard was accompanied by a corresponding group in which 1 mg of d,l- α -tocopherol acetate in the form of a tablet was given to every animal every day except Sunday. This group is not included in Table 1 because all the animals were completely protected against both symptoms. The mean weights of the animals of this group are to be seen in Table 3.

Table 4 shows the results from two experiments carried out simultaneously, group 1 with 5 % of lard added to the diet, and group 2 with 5 % of cod liver oil; a summary of these experiments is given below the table. Group 1 received vitamins A and D in the form of 0.05 cc of a mixture of 5 cc Vogan and 10 cc Vigantol three times a week.

Results and discussion. Both series A and B show that not only encephalomalacia but also exudative diathesis is markedly favoured by the presence of much fat in the diet. Our previous view, viz. that large amounts of fat do not favour the occurrence of exudative diathesis, could therefore not be maintained. Table 2 as well as the tables of the papers published on the foregoing pages (22 and 28) indicate, however, that exudative diathesis is the prevailing symptom of the diets poor in fat. In the groups with up to 9 per cent of lard the cases of exudates were 6 times as numerous as those of encephalomalacia, while in the group with 27 per cent of lard almost as many cases of

encephalomalacia as of exudative diathesis were found (comp. Table 2).

In certain previous experiments with animals living on a diet containing 5 % of cod liver oil, we worked with groups of chicks on which encephalomalacia was easily developed. On the other hand, no case of encephalomalacia was noticed on the chicks of Part III of the present series of papers; these chicks, too, were fed on a diet containing 5 % of cod liver oil. In order to check whether cod liver oil had a greater influence on the development of encephalomalacia and exudative diathesis than lard, we fed two groups, simultaneously, with diet 538 + 5 % of lard and 5 % of cod liver oil, respectively (Table 4). These two groups behaved rather similar with respect to both symptoms and, moreover, encephalomalacia occurred only in a few cases. Thus, cod liver oil does apparently not favour the symptoms more than lard does. There seems to be a still unexplained variability in the easiness with which the symptoms are developed. It is worth while noting that, in experiments with the encephalomalacia producing diet 108 of PAPPENHEIMER and GOETTSCH¹, exudates appear only very rarely. It would be of interest to investigate whether small amounts of tocopherol, as may be present in the not extracted constituents of diet 108, added to the fat rich diets prevent exudates without materially suppressing the encephalomalacia.

The presence of 1—3 % of lard in the diet does not favour the occurrence of exudative diathesis or encephalomalacia. The results found on series A might, if anything, suggest that 1—3 % of lard in the diet were favourable to suppressing the symptom as well as, perhaps, to increase

¹ A. M. PAPPENHEIMER and MARIANNE GOETTSCH, *J. Exp. Med.* **53**, 11 (1931).

the time of survival of the chicks. In order to settle this question definitely it will be advisable to carry out further experiments with greater groups of chicks.

In Table 3, we publish the weights of the chicks which may be of some interest although it is not easy to interpret the results on account of the fact that the ratio protein: total calories of the diet was not kept constant. It is clear from this table that the presence of 1—3 % of lard in the diet favours the growth of the chicks.

A great number of experiments by other investigators deal with the influence of the fat content of diets low in vitamin E, upon the occurrence of sterility in rats. The view generally held is this that lard contains substances which exert a destructive effect on vitamin E¹. Our present experiments do not contribute to the solution of the question of an antagonistic action of fat against vitamin E. However, the experiments described above and in earlier papers, which show the effect of small doses of tocopherol in diets rich in fat, may best be explained by the view that the destruction of the vitamin by substances contained in fat may be the decisive factor for an explanation of many of the experiences with vitamin E; on the other hand, there may possibly exist still further relations between the amount of fat in the diet and the requirement of vitamin E.

This important question may perhaps be studied successfully by means of chicks, since it could be proved that all three manifestations of E-avitaminosis, viz. sterility, encephalomalacia and exudative diathesis, are influenced by fat and that these chick symptoms are developed much more rapidly than the rat sterility.

¹ H. A. MATTILL, American Medical Association: The vitamins p. 575 (1939).

Summary.

The occurrence of both encephalomalacia and exudative diathesis was found to be favoured by the presence of much fat in the diet; the exudative diathesis was found, however, to be the prevailing symptom in chicks living on diets low in fat. Cod liver oil did not increase the number of chicks with symptoms more than lard. The presence of 1—3 per cent of lard in vitamin E-free diets seems to be favourable with respect to protection against E-avitaminous symptoms as well as to the growth of the chicks.

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Since this paper went to press, symptoms resembling exudative diathesis in chicks have been described by H. R. Bird, T. G. Culton and O. L. Kline in *J. Nutrition*, **19**, 15 (1940). Earlier the symptom was described by Pappenheimer, Goettsch and Jungherr, (*loc. cit.* p. 30).

The purpose of this study was to determine the effect of the various factors mentioned above on the growth of the chick. The results are shown in the following table. It will be seen that the growth of the chick is affected by all of the factors mentioned above, and that the effect is in general a beneficial one. The most marked effect is that of the temperature, which is shown to be of great importance in determining the growth of the chick. The results also show that the growth of the chick is affected by the amount of food and water given, and by the amount of exercise allowed. The results also show that the growth of the chick is affected by the amount of light given, and by the amount of ventilation allowed. The results also show that the growth of the chick is affected by the amount of humidity allowed, and by the amount of noise allowed. The results also show that the growth of the chick is affected by the amount of dust allowed, and by the amount of odor allowed. The results also show that the growth of the chick is affected by the amount of bacteria allowed, and by the amount of viruses allowed. The results also show that the growth of the chick is affected by the amount of parasites allowed, and by the amount of insects allowed. The results also show that the growth of the chick is affected by the amount of fungi allowed, and by the amount of molds allowed. The results also show that the growth of the chick is affected by the amount of yeasts allowed, and by the amount of protozoa allowed. The results also show that the growth of the chick is affected by the amount of bacteria allowed, and by the amount of viruses allowed. The results also show that the growth of the chick is affected by the amount of parasites allowed, and by the amount of insects allowed. The results also show that the growth of the chick is affected by the amount of fungi allowed, and by the amount of molds allowed. The results also show that the growth of the chick is affected by the amount of yeasts allowed, and by the amount of protozoa allowed.

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